Multiplex HS MasterMix (2X)

with antibody inhibited Taq DNA polymerase

| Product | Cat# | Package size |
|---|------------|--------------|
| Premixed PCR master mix optimized for Multiplex-PCR. Mix for 100x20µL reaction volume. | M3013.0100 | 1mL |
| Premixed PCR master mix optimized for Multiplex-PCR. Mix for 500x20µL reaction volume. | M3013.0500 | 5 x 1mL |

Product Description

Multiplex PCR is a method that enables amplification of two or more amplicons simultaneously in a single reaction tube/reaction. It is widely used in genotyping and different areas of DNA testing in research, forensic and diagnostic laboratories.

The Genaxxon bioscience Multiplex HS Mastermix (2X) facilitates the amplification of multiple PCR products, minimizes the need for optimization, making the development of multiplex PCR assays both fast and simple.

Our Multiplex HS Mastermix (2X) is a ready-to-use PCR mixture is an optimized ready-to-use mixture for probe-based assays such as TaqMan®, Beacons and MGBs. It contains a modified fast HotStart Taq DNA Polymerase, dNTPs and MgCl₂ combined in an optimized buffer system for Real-Time PCR / qPCR applications except primers, probe and template DNA / cDNA.

The HotStart Taq Polymerase is based on the standard *Taq* DNA polymerase from Genaxxon inactivated by a specific antibody against *Taq* DNA polymerase which is activated by heat treatment. Thus, during setup and the first ramp of thermal cycling, the enzyme is not active and misprimed primers are not extended. The result is higher specificity, increased sensitivity and greater yields when compared to standard DNA polymerases, making this enzyme especially well-suited for multiplex PCR.

Product Specifications

| Concentration: | 2 time ready-to-use master mix |
|-----------------------------|--------------------------------|
| Extension rate: | 4-6 kb/min. at 72°C |
| Half-life: | 10min. at 97°C, 60min. at 94°C |
| 5'-3' exonuclease activity: | Yes |
| Extra addition of A: | Yes |
| 3'-5' exonuclease activity: | No |
| Nuclease contamination: | No |
| Protease contamination: | No |
| RNase contamination: | No |
| Self-priming activity: | No |

Unit definition

One unit of HotStart Taq DNA-Polymerase used for the Genaxxon bioscience Multiplex HS Mastermix (2X) is defined as the amount of enzyme that incorporates 10nmol of dNTP's into acid-insoluble fraction in 30 minutes at 72°C under standard assay conditions.

Quality Control

| Amplification efficiency: | Amplification efficiency is tested in parallel amplification reactions and additionally against competitors products. |
|---------------------------|---|
| PCR reproducibility: | PCR reproducibility is tested in parallel amplification reaction. |
| Exonuclease activity: | Linearized DNA is incubated with HotStart Tag DNA polymerase in Multiplex HS PCR buffer. |
| Endonuclease activity: | Plasmid DNA is incubated with HotStart Tag DNA polymerase in Multiplex HS PCR buffer. |
| RNase activity: | RNA is incubated with HotStart Tag DNA polymerase in Multiplex HS PCR buffer. |
| Protease activity: | HotSTart Tag DNA polymerase is incubated in storage buffer. |
| Self-priming activity: | PCR is performed under standard conditions, without primers, using HotStart Taq DNA polymerase and human genomic DNA. |

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Application

Automated hotstart PCR Gene expression Multiplex qPCR / Multiplex PCR with fluorochromes Genotyping Copy number analysis

Stability

The Genaxxon bioscience Multiplex HS Mastermix is shipped on wet ice but retain full activity at RT (+15 - +25°C) for at least 3-4 days.

The Genaxxon bioscience Multiplex HS Mastermix, including buffers and reagents, should be stored immediately upon receipt at -20°C. When stored under these conditions and handled correctly, these products can be kept at least until the expiration date (see tube label) without showing any reduction in performance.

The Genaxxon bioscience Multiplex HS Mastermix can also be stored at +2°C to +8°C°C up to 3 months.

Product Use Limitations

The Multiplex HS Mastermix 2X is developed, designed, and sold for research purposes only. It is not to be used for human, diagnostic or drug purposes or to be administered to humans unless expressly cleared for that purpose by the Food and Drug Administration in the USA or the appropriate regulatory authorities in the country of use. All due care and attention should be exercised in the handling of many of the materials described in this manual.

Safety information

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, please consult the appropriate material safety data sheets (MSDSs). These are available online as pdf-file or on request (info@genaxxon.com).

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PCR Protocol Part

Protocol using Multiplex HS Mastermix (2X)

This protocol serves as a guideline for PCR amplification. Optimal reaction conditions such as incubation times, temperatures, and amount of template DNA may vary and must be determined individually.

Important notes before getting started

- All primers should have the same melting temperature.
- Use equal concentrations of all primers for the initial test.
- If one product gives a much stronger band than the others, reduce the primer concentration for this target.
- If one product gives a much weaker band than the others, increase the primer concentration of this target.
- Set up all reaction mixtures in an area separate from that used for DNA preparation or PCR product analysis. Working on ice is not required.
- The MgCl2 concentration in the final reaction is 3mM. In some applications, more MgCl2 is required for best results.
- Use disposable tips containing hydrophobic filters to minimize cross-contamination.

Procedure

1. Thaw primer solutions

Keep on ice after complete thawing, and mix well before use.

Optional: Prepare a primer mix of an appropriate concentration using nuclease-free water. This is recommended if several amplification reactions using the same primer pair are to be performed. The final volume of diluted primer mix plus the template DNA, added at step 4, should not exceed 12.5μ L per reaction.

2. Thaw Multiplex HS Mastermix (2X) at RT or on ice.

It is very important to mix the Multiplex HS Mastermix well before use to avoid local differences in salt concentration. Spin vials briefly after mixing. The Genaxxon bioscience Multiplex HS Mastermix is provided as a 2X concentrated (i.e., a 10µL volume of Multiplex HS Mastermix is required for PCR reactions with a final volume of 20µL). For volumes smaller than 20µL, the 1:1 ratio of Multiplex HS Mastermix to diluted primer mix, template DNA and water should be maintained. A negative control (PCR without template DNA) should be included in every experiment.

- 3. Distribute the appropriate volume of diluted primer mix into the PCR tubes containing the Multiplex HS Mastermix.
- 4. Add template DNA to the individual PCR tubes containing the reaction mix.

 Table 1: PCR reaction components using Multiplex HS Mastermix (2X) (20µL PCR reaction)

| Components | Quantities/Volumes | Final concentrations/amount |
|--|--|---|
| Multiplex HS Mastermix: each forward primer (10µM): each reverse primer (10µM): Probe Template DNA | 10μL variable, e.g. 2μL variable, e.g. 2μL variable, e.g. 2μL variable | 1x 0.1 - 0.5μM 0.1 - 0.5μM 0.2 - 0.5μM genomic DNA: 1 - 20ng plasmid DNA: 1 - 50pg bacterial DNA: 0.1 - 1ng |
| Nuclease-free water | adjust to 20µL | |
| Total volume | 20µL | |

The table above shows the suggested starting conditions. Please adopt the protocol to your needs. Theoretically usable conditions in brackets.

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- 1. **Program the thermal cycler** according to the manufacturer's instructions. A typical PCR cycling program is outlined in Table 3. For maximum yield and specificity, temperatures and cycling times should be optimized for each new target or primer pair.
- 2. Place PCR tubes in the thermal cycler and start program.

Table 2: PCR conditions (Thermal cycler) - 3-step PCR protocol

| Step | time | temperature | comments |
|-----------------------|----------------|-------------|--|
| Initial denaturation: | 2-3 min. | 95°C | |
| Denaturation: | 5 - 10 seconds | 95°C | |
| Annealing: | 5 - 10 seconds | 50 - 68°C | Approximately 3 - 5°C below Tm of primers. |
| Extension: | 5 - 10 seconds | 72°C | For PCR products longer than 1kb, use an extension time of approximately 1min./kb DNA. |
| Number of Cycles | 30 - 45 | | |
| Final extension | 5 min. | 72°C | Occasionally used |

Table 3: PCR conditions (Thermal cycler) - 2-step PCR protocol

| Step | time | temperature | comments |
|------------------------------|----------------|-------------|---|
| Initial denaturation: | 2-3 min. | 95°C | |
| Denaturation: | 5 - 10 seconds | 95°C | |
| Annealing/Extension combined | 5 - 10 seconds | 60 - 68°C | Depends on primers. Approximately 3 - 5°C below Tm of primers. |
| Number of Cycles | 30 - 45 | | |
| Final extension | 5 min. | 72°C | Occasionally used |

Note: After amplification, samples can be stored at $+2^{\circ}C$ to $+8^{\circ}C$ overnight, or $-20^{\circ}C$ for long term storage.

Note: For maximum efficiency and specificity, annealing temperatures, as well as extension time, primer/probe concentration and template DNA concentration may need to be optimized.

Table 4: Migration Chart of some Gel Tracking Dyes

| Dye in agarose gel | 0.5%-1.5% | 2.0%-3.0% | CAS- number | Cat-No. Genaxxon |
|-----------------------|------------------|---------------|----------------|---------------------|
| Xylene cyanol | 10000bp - 4000bp | 750bp - 200bp | 2650-17-1 | M3312 |
| Cresol Red | 2000bp - 1000bp | 200bp - 125bp | 62625-29-0 | M3371 |
| Bromophenol blue | 500bp - 400bp | 150bp - 50bp | 115-39-9 | M3092 |
| Orange G | <100bp | <20bp | 1936-15-8 | M3180 |
| Tartrazine | <20bp | <20bp | 1934-21-0 | |

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